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ACTIVE IMMUNIZATION TO GENERATE ANTIBODIES TO SOLUBLE A-BETA AUG 2005

CROSS-REFERENCES TO RELATED APPLICATIONS

[0001] This application is an application claiming benefit under 35 U.S.C. § 119(e) of U.S. Application No. 60/444,150, filed February 1, 2003, which is incorporated by reference in its entirety for all purposes.

TECHNICAL FIELD

[0002] The invention resides in the technical fields of immunology and medicine.

BACKGROUND OF THE INVENTION

[0003] Alzheimer's disease (AD) is a progressive disease resulting in senile dementia. See generally Selkoe, TINS 16, 403-409 (1993); Hardy et al., WO 92/13069; Selkoe, J. Neuropathol. Exp. Neurol. 53, 438-447 (1994); Duff et al., Nature 373, 476-477 (1995); Games et al., Nature 373, 523 (1995). Broadly speaking, the disease falls into two categories: late onset, which occurs in old age (65 + years) and early onset, which develops well before the senile period, i.e., between 35 and 60 years. In both types of disease, the pathology is the same but the abnormalities tend to be more severe and widespread in cases beginning at an earlier age. The disease is characterized by at least two types of lesions in the brain, senile plaques and neurofibrillary tangles. Senile plaques are areas of disorganized, neuropil up to 150 µm across with extracellular amyloid deposits at the center visible by microscopic analysis of sections of brain tissue. Neurofibrillary tangles are intracellular deposits of microtubule associated tau protein consisting of two filaments twisted about each other in pairs.

[0004] The principal constituent of the plaques is a peptide termed Aβ or β-amyloid peptide. Aβ peptide is an internal fragment of 39-43 amino acids of a precursor protein termed amyloid precursor protein (APP). Several mutations within the APP protein have been correlated with the presence of Alzheimer's disease. See, e.g., Goate et al., Nature 349, 704) (1991) (valine⁷¹⁷ to isoleucine); Chartier Harlan et al. Nature 353, 844 (1991)) (valine⁷¹⁷ to glycine); Murrell et al., Science 254, 97 (1991) (valine⁷¹⁷ to phenylalanine); Mullan et al., Nature Genet. 1, 345 (1992) (a double mutation changing lysine⁵⁹⁵-methionine⁵⁹⁶ to asparagine⁵⁹⁵-leucine⁵⁹⁶). Such mutations are thought to cause Alzheimer's disease by

increased or altered processing of APP to A β , particularly processing of APP to increased amounts of the long form of A β (i.e., A β 1-42 and A β 1-43). Mutations in other genes, such as the presentiin genes, PS1 and PS2, are thought indirectly to affect processing of APP to generate increased amounts of long form A β (see Hardy, TINS 20, 154 (1997)). These observations indicate that A β , and particularly its long form, is a causative element in Alzheimer's disease.

[0005] Immunization of transgenic mouse models of AD with β-amyloid peptide (Aβ) derived-immunogens results in an antibody response that inhibits formation and/or clear amyloid plaques in brains of the mice (Schenk et al., (1999) Nature 400, 173-177; Janus et al., (2000) Nature 408, 979-982, Morgan et al. (2000) Nature 408, 982-985, Sigurdsson et al., (2001) Am. J. Pathol. 159, 439-447.1-4)). Passively administered antibodies to Aß have achieve similar effects. Antibody-mediated, Fc-dependent phagocytosis by microglial cells and/or macrophages has been proposed as one mechanism for clearance of existing amyloid plaques (Bard et al., (2000) Nat. Med., 6, 916-919)). This proposal is based on the result that certain peripherally administered antibodies against AB enter the CNS of transgenic mice, decorate amyloid plaques, and induce plaque clearance. Also, a strong correlation has been reported between antibodies that were efficacious in vivo and in an ex vivo assay using sections of PDAPP or Alzheimer's disease (AD) brain to measure plaque clearing activity. Fc receptors on microglial cells effected the clearance response in the ex vivo assay. However, it has been also been reported that antibody efficacy can also be obtained in vivo by mechanisms that are independent of Fc interactions (Bacskai et al., (2002) J. Neurosci., 22, 7873-7878). An antibody directed against the mid-portion of Aβ, which cannot recognize amyloid plaques, was reported to bind to soluble AB and reduce plaque deposition (DeMattos et al., (2001) Proc. Natl. Acad. Sci. USA, 98, 8850-8855). Short-term treatment with this antibody has also been reported to improve performance in an object recognition task without affecting amyloid burden (Dodart et al., (2002) Nat. Neurosci., 5, 452-457).

[0006] This application is related to WO 00/72880 filed May 26, 2000, WO 99/27944, filed November 30, 1998, U.S. Application No. 60/067,740, filed December 2, 1997, U.S. Application No. 60/080,970, filed April 7, 1998, and U.S. Application No. 09/201,430, filed November 30, 1998, each of which is incorporated by reference in its entirety for all purposes.

BRIEF DESCRIPTION OF THE DRAWINGS

[0007] Figs. 1A-C. Antibodies produced by immunization with N-terminal fragments of A β bind to amyloid plaques. Fig. 1A. Peptides encompassing various domains of A β 1-42 (SEQ ID NO:1) (synthesized contiguous to T cell epitope derived from ovalbumin) were used to immunize PDAPP mice. A reversemer, A β 5-1 (SEQ ID NO:2), was used as a negative control. Fig. 1B. ELISA titers against aggregated A β 1-42 were significantly higher over the length of the study in the A β 5-11 and A β 15-24 groups than in the A β 1-5 group (1:14,457, p<0.01 and 1:12,257, p<0.05 vs. 1:3,647, respectively; ANOVA followed by *post hoc* Tukey's test). Fig. 1C. Unfixed cryostat sections from untreated PDAPP mouse brain were exposed to the sera of mice immunized with A β 5-1, A β 3-9, A β 5-11, or A β 15-24 (titers normalized to 1:1000 for staining). Antibodies to A β 15-24 did not bind to amyloid plaques. Scale bar represents 500 μ m.

[0008] Figs. 2A-C. Capture of soluble Aβ1-42 by antibodies is not associated with reduced amyloid burden or neuritic pathology. Fig. 2A. Sera from mice immunized with fragments of A β were examined for their ability to capture radiolabeled soluble A β 1-42 in a radioimmunoassay. Sera from all animals immunized with A\beta15-24 were able to capture soluble A\beta 1-42 (one serum sample had a titer higher than 1:1,350 and a precise titer was not determined), compared with 27% of those in the A\u00e31-5 group and 3% of the A\u00e33-9 group. Figs.2B-C. Amyloid burden (Fig. 2B) and neuritic pathology (Fig. 2C) were evaluated with image analysis by a blinded microscopist. Values are expressed as a percentage of the mean of the A β 5-1 group (negative control reversemer peptide). The A β 5-11 group was evaluated at a separate sitting from the other groups, but in conjunction with the same negative control group as an internal reference (second Aβ5-1 reversemer set, on the left). Amyloid burden was significantly reduced in the Aβ1-5, Aβ3-9, and Aβ5-11 groups (p<0.001. Bars represent median values and the dashed horizontal line indicates the control level. Neuritic burden was significantly reduced in the Aβ3-9 and Aβ5-11 groups (p<0.05). Neither endpoint was significantly altered by immunization with AB15-24 group. Statistical analysis was preformed with square root transformation (to normalize non-parametric distributions), and analyzed with ANOVA. A Dunnett's test was then used to compare the multiple groups

A β 1-5, A β 3-9, A β 15-24 groups with their A β 5-1 control, and Mann-Whitney for the A β 5-11 group with its corresponding A β 5-1 reversemer control.

DETAILED DESCRIPTION

I. General

[0009] The invention provides methods of preventing, effecting prophylaxis of, or treating a disease associated with amyloid deposits using fragments from a central or C-terminal regions of A β . Such fragments can induce a polyclonal mixture of antibodies that specifically bind to soluble A β without binding to plaques. The antibodies can inhibit formation of amyloid deposits of A β in the brain of a patient from soluble A β , thus preventing or treating the disease. Fragment A β 15-24 and subfragments of 5-10 contiguous amino acids thereof are preferred immunogens due to their capacity to generate a high titer of antibodies.

II. <u>DEFINITIONS</u>

[0010] For purposes of classifying amino acids substitutions as conservative or nonconservative, amino acids are grouped as follows: Group I (hydrophobic sidechains): norleucine, met, ala, val, leu, ile; Group II (neutral hydrophilic side chains): cys, ser, thr; Group III (acidic side chains): asp, glu; Group IV (basic side chains): asn, gln, his, lys, arg; Group V (residues influencing chain orientation): gly, pro; and Group VI (aromatic side chains): trp, tyr, phe. Conservative substitutions involve substitutions between amino acids in the same class. Non-conservative substitutions constitute exchanging a member of one of these classes for a member of another.

[0011] The term "all-D" refers to peptides having $\geq 75\%$, $\geq 80\%$, $\geq 85\%$, $\geq 90\%$, $\geq 95\%$, and 100% D-configuration amino acids.

[0012] The term "agent" is used to describe a compound that has or may have a pharmacological activity. Agents include compounds that are known drugs, compounds for which pharmacological activity has been identified but which are undergoing further therapeutic evaluation, and compounds that are members of collections and libraries that are to be screened for a pharmacological activity.

[0013] Therapeutic agents of the invention are typically substantially pure from undesired contaminant. This means that an agent is typically at least about 50% w/w (weight/weight) purity, as well as being substantially free from interfering proteins and contaminants.

Sometimes the agents are at least about 80% w/w and, more preferably at least 90 or about

95% w/w purity. However, using conventional protein purification techniques, homogeneous peptides of at least 99% w/w can be obtained. Therapeutic agents of the invention may prevent, effect prophylaxis of, or treat a disease associated with amyloid deposits.

[0014] Specific binding between two entities means the entities have a mutual affinity for each other that is at least 10-, 100- or 100-fold greater than the affinity of either entity for a control, such as unrelated antigen or antibody to a different antigen. The mutual affinity of the two entities for each other is usually at least 10^7 , 10^8 , 10^9 M⁻¹, or 10^{10} M⁻¹. Affinities greater than 10^8 M⁻¹ are preferred. Specific binding of a polyclonal antibody to an epitope within A β means the antibodies in the polyclonal antibody population specifically bind to one epitope of A β without binding to other epitopes of A β .

[0015] The term "antibody" or "immunoglobulin" is used to include intact antibodies and binding fragments thereof. Typically, fragments compete with the intact antibody from which they were derived for specific binding to an antigen fragment including separate heavy chains, light chains Fab, Fab' F(ab')2, Fabc, and Fv. Fragments are produced by recombinant DNA techniques, or by enzymatic or chemical separation of intact immunoglobulins. The term "antibody" also includes one or more immunoglobulin chains that are chemically conjugated to, or expressed as, fusion proteins with other proteins. The term "antibody" also includes bispecific antibody. A bispecific or bifunctional antibody is an artificial hybrid antibody having two different heavy/light chain pairs and two different binding sites. Bispecific antibodies can be produced by a variety of methods including fusion of hybridomas or linking of Fab' fragments. See, e.g., Songsivilai & Lachmann, Clin. Exp. Immunol. 79:315-321 (1990); Kostelny et al., J. Immunol. 148, 1547-1553 (1992).

[0016] A β , also known as β -amyloid peptide, or A4 peptide (see US 4,666,829; Glenner & Wong, *Biochem. Biophys. Res. Commun.*, 120, 1131 (1984)), is a peptide of 39-43 amino acids, which is the principal component of characteristic plaques of Alzheimer's disease. A β has several natural occurring forms. The natural human forms of A β are referred to as A β 39, A β 40, A β 41, A β 42 and A β 43. The sequences of these peptides and their relationship to the APP precursor are illustrated by Fig. 1 of Hardy et al., TINS 20, 155-158 (1997). For example, A β 42 has the sequence:

[0017] H₂N-Asp-Ala-Glu-Phe-Arg-His-Asp-Ser-Gly-Tyr-Glu-Val-His-His-Gln-Lys-Leu-Val-Phe-Phe-Ala-Glu-Asp-Val-Gly-Ser-Asn-Lys-Gly-Ala-Ile-Ile-Gly-Leu-Met-Val-Gly-Gly-Val-Val-Ile-Ala-OH (SEQ ID NO:1).

[0018] A β 41, A β 40 and A β 39 differ from A β 42 by the omission of Ala, Ala-Ile, and Ala-Ile-Val respectively from the C-terminal end. A β 43 differs from A β 42 by the presence of a threonine residue at the C-terminus.

[0019] APP⁶⁹⁵, APP⁷⁵¹, and APP⁷⁷⁰ refer, respectively, to the 695, 751, and 770 amino acid residue long polypeptides encoded by the human APP gene. See Kang et al., *Nature*, 325, 773 (1987); Ponte et al., *Nature*, 331, 525 (1988); and Kitaguchi et al., *Nature*, 331, 530 (1988). Amino acids within the human amyloid precursor protein (APP) are assigned numbers according to the sequence of the APP770 isoform. Terms such as Aβ39, Aβ40, Aβ41, Aβ42 and Aβ43 refer to an Aβ peptide containing amino acid residues 1-39, 1-40, 1-41, 1-42 and 1-43, respectively.

[0020] Disaggregated A β or fragments thereof means monomeric peptide units. Disaggregated A β or fragments thereof are generally soluble, and are capable of self-aggregating to form soluble oligomers. Oligomers of A β and fragments thereof are usually soluble and exist predominantly as alpha-helices or random coils. One method to prepare monomeric A β is to dissolve lyophilized peptide in neat DMSO with sonication. The resulting solution is centrifuged to remove any insoluble particulates. Aggregated A β or fragments thereof, means oligomers of A β or immunogenic fragments thereof in which the monomeric units are held together by noncovalent bonds and associate into insoluble beta-sheet assemblies. Aggregated A β or fragments thereof, means also means fibrillar polymers. Fibrils are usually insoluble. Some antibodies bind either soluble A β or fragments thereof or aggregated A β or fragments thereof. Some antibodies bind both soluble A β or fragments thereof and aggregated A β or fragments thereof. Some antibodies bind to soluble A β without binding to plaque.

[0021] An "antigen" is an entity to which an antibody specifically binds.

[0022] The term "epitope" or "antigenic determinant" refers to a site on an antigen to which B and/or T cells respond. B-cell epitopes can be formed both from contiguous amino acids or noncontiguous amino acids juxtaposed by tertiary folding of a protein. Epitopes formed from contiguous amino acids are typically retained on exposure to denaturing solvents whereas epitopes formed by tertiary folding are typically lost on treatment with denaturing solvents. An epitope typically includes at least 3, and more usually, at least 5 or 8-10 amino acids in a unique spatial conformation. Methods of determining spatial conformation of epitopes include, for example, x-ray crystallography and 2-dimensional nuclear magnetic resonance.

See, e.g., Epitope Mapping Protocols in Methods in Molecular Biology, Vol. 66, Glenn E. Morris, Ed. (1996). Antibodies that recognize the same epitope can be identified in a simple immunoassay showing the ability of one antibody to block the binding of another antibody to a target antigen. T-cells recognize continuous epitopes of about nine amino acids for CD8 cells or about 13-15 amino acids for CD4 cells. T cells that recognize the epitope can be identified by in vitro assays that measure antigen-dependent proliferation, as determined by ³H-thymidine incorporation by primed T cells in response to an epitope (Burke et al., J. Inf. Dis., 170, 1110-19 (1994)), by antigen-dependent killing (cytotoxic T lymphocyte assay, Tigges et al., J. Immunol., 156, 3901-3910) or by cytokine secretion.

[0023] An N-terminal epitope of $A\beta$ means an epitope with residues 1-11. An epitope within a C-terminal region means an epitope within residues 29-43, and an epitope within a central regions means an epitope with residues 12-28.

[0024] The term "immunological" or "immune" response is the development of a beneficial humoral (antibody mediated) and/or a cellular (mediated by antigen-specific T cells or their secretion products) response directed against an amyloid peptide in a recipient patient. Such a response can be an active response induced by administration of immunogen or a passive response induced by administration of antibody or primed T-cells. A cellular immune response is elicited by the presentation of polypeptide epitopes in association with Class I or Class II MHC molecules to activate antigen-specific CD4⁺ T helper cells and/or CD8⁺ cytotoxic T cells. The response may also involve activation of monocytes, macrophages, NK cells, basophils, dendritic cells, astrocytes, microglia cells, eosinophils or other components of innate immunity. The presence of a cell-mediated immunological response can be determined by proliferation assays (CD4⁺ T cells) or CTL (cytotoxic T lymphocyte) assays (see Burke, *supra*; Tigges, *supra*). The relative contributions of humoral and cellular responses to the protective or therapeutic effect of an immunogen can be distinguished by separately isolating antibodies and T-cells from an immunized syngeneic animal and measuring protective or therapeutic effect in a second subject.

[0025] An "immunogenic agent" or "immunogen" is capable of inducing an immunological response against itself on administration to a mammal, optionally in conjunction with an adjuvant.

[0026] The term "naked polynucleotide" refers to a polynucleotide not complexed with colloidal materials. Naked polynucleotides are sometimes cloned in a plasmid vector.

[0027] The term "adjuvant" refers to a compound that when administered in conjunction with an antigen augments the immune response to the antigen, but when administered alone does not generate an immune response to the antigen. Adjuvants can augment an immune response by several mechanisms including lymphocyte recruitment, stimulation of B and/or T cells, and stimulation of macrophages.

[0028] The term "patient" includes human and other mammalian subjects that receive either prophylactic or therapeutic treatment.

[0029] Competition between antibodies is determined by an assay in which the immunoglobulin under test inhibits specific binding of a reference antibody to a common antigen, such as Aβ. Numerous types of competitive binding assays are known, for example: solid phase direct or indirect radioimmunoassay (RIA), solid phase direct or indirect enzyme immunoassay (EIA), sandwich competition assay (see Stahli et al., Methods in Enzymology, 9:242-253 (1983)); solid phase direct biotin-avidin EIA (see Kirkland et al., J. Immunol. 137:3614-3619 (1986)); solid phase direct labeled assay, solid phase direct labeled sandwich assay (see Harlow and Lane, "Antibodies, A Laboratory Manual," Cold Spring Harbor Press (1988)); solid phase direct label RIA using I-125 label (see Morel et al., Molec, Immunol. 25(1):7-15 (1988)); solid phase direct biotin-avidin EIA (Cheung et al., Virology, 176:546-552 (1990)); and direct labeled RIA (Moldenhauer et al., Scand. J. Immunol., 32:77-82 (1990)). Typically, such an assay involves the use of purified antigen bound to a solid surface or cells bearing either of these, an unlabelled test immunoglobulin and a labeled reference immunoglobulin. Competitive inhibition is measured by determining the amount of label bound to the solid surface or cells in the presence of the test immunoglobulin. Usually the test immunoglobulin is present in excess. Antibodies identified by competition assay (competing antibodies) include antibodies binding to the same epitope as the reference antibody and antibodies binding to an adjacent epitope sufficiently proximal to the epitope bound by the reference antibody for steric hindrance to occur. Usually, when a competing antibody is present in excess, it will inhibit specific binding of a reference antibody to a common antigen by at least 50 or 75%.

[0030] An antibody that specifically binds to soluble A β means an antibody that binds to soluble A β with an affinity of at least 10^7 M^{-1} . Some antibodies bind to soluble A β with affinities between 10^8 M^{-1} and 10^{11} M^{-1} .

[0031] An antibody that specifically binds to soluble A β without specifically binding to plaques means an antibody that binds to soluble A β as described above and has at least a ten fold and usually at least 100-fold lower specific binding affinity for plaques (*i.e.*, A β in aggregated β -pleated sheet form) from a cadaver of a former Alzheimer's patient or a transgenic animal model. For example, such an antibody might bind to soluble A β with an affinity of 10^9 M⁻¹ and to plaques with an affinity less than 10^7 M⁻¹. The affinity of such antibodies for plaques is usually less than 10^7 or 10^6 M⁻¹. Such antibodies are additionally or alternatively defined by fluorescence intensity relative to an irrelevant control antibody (*e.g.*, an antibody or mixture of polyclonal antibodies to a reversemer A β peptide) when the antibodies are contacted with plaques and binding assessed by fluorescently labeling (as described in the Examples section). The fluorescence intensity of antibodies that bind to soluble A β peptide without binding to plaques is within a factor of five, sometimes within a factor of two and sometimes indistinguishable within experimental error from that of the control antibody.

[0032] Compositions or methods "comprising" one or more recited elements may include other elements not specifically recited. For example, a composition that comprises $A\beta$ peptide encompasses both an isolated $A\beta$ peptide and $A\beta$ peptide as a component of a larger polypeptide sequence.

III. Aß peptides for Active Immunization

[0033] A β peptides for use in the methods of the invention are immunogenic peptides that on administration to a human patient or animal generate antibodies that specifically bind to one or more epitopes between residues 12 and 43 of A β without generating antibodies that specifically bind to one or more epitopes within residues 1-11 of A β . Antibodies specifically binding to epitopes between residues 12 and 43 specifically bind to soluble A β without binding to plaques of A β . These types of antibody can specifically bind to soluble A β in the circulation of a patient or model amyloid without specifically binding to plaques of A β deposits in the brain of the patient or model. The specifically binding of antibodies to soluble A β inhibits the A β from being incorporated into plaques thus either inhibiting development of the plaques in a patient or inhibiting a further increase in the size or frequency of plaques if such plaques have already developed before treatment is administered.

[0034] Preferably, the fragment of Aβ administered lacks an epitope that would generate a T-cell response to the fragment. Generally, T-cell epitopes are greater than 10 contiguous amino acids. Therefore, preferred fragments of Aβ are of size 5-10 or preferably 7-10 contiguous amino acids; *i.e.*, sufficient length to generate an antibody response without generating a T-cell response. Absence of T-cell epitopes is preferred because these epitopes are not needed for immunogenic activity of fragments, and may cause an undesired inflammatory response in a subset of patients (Anderson et al., (2002) *J. Immunol.* 168, 3697-3701; Senior (2002) Lancet *Neurol.* 1, 3). In some methods, the fragment is a fragment other than Aβ13-28, 17-28, 25-35, 35-40, 33-42 or 35-42. Most T-cell epitopes occur within amino acids 14-30 of Aβ.

[0035] Fragment Aβ15-24 and subfragments of 7-9 contiguous amino acids thereof are preferred because these peptides consistently generate a high immunogenic response to Aβ peptide. These fragments include Aβ15-21, Aβ16-22, Aβ17-23, Aβ18-24, Aβ19-25, Aβ15-22, Aβ16-23, Aβ17-24, Aβ18-25, Aβ15-23, Aβ16-24, Aβ17-25, Aβ18-26, Aβ15-24, Aβ16-25, and Aβ15-25. The designation Aβ15-21 for example, indicates a fragment including residues 15-21 of Aβ and lacking other residues of Aβ. Also preferred are C-terminal fragments of Aβ42 or 43 of 5-10 and preferably 7-10 contiguous amino acids. These fragments can generate an antibody response that includes end-specific antibodies. These antibodies are advantageous in specifically binding to Aβ42 and Aβ43 without specifically binding to Aβ39-41. These antibodies bind to soluble Aβ without binding to plaque.

[0036] In some methods, a fragment from the central or C-terminal region of A β is administered in a regime that also includes administering a fragment from the N-terminal region. In general, such fragments induce antibodies that specifically bind to and induce clearing of amyloid plaques via phagocytic cells. Such a response is particularly useful to clear existing deposits of A β . However, once the deposits have been cleared, further treatment with a fragment from the central or C-terminal region of A β to induce antibodies to soluble A β is advantageous for preventing further deposition of A β without risk of inflammatory side effects in certain patients. N-terminal fragments beginning at residues 1-3 of A β and ending at residues 7-11 of A β are particularly preferred. Exemplary N-terminal fragments include A β 1-5, 1-6, 1-7, 1-10, 3-7, 1-3, and 1-4.

[0037] Unless otherwise indicated, reference to A β includes the natural human amino acid sequences indicated above as well as analogs including allelic, species and induced variants. Analogs of A β induce antibodies that specifically bind with a natural A β peptide (e.g., A β 42). Analogs of A β typically differ from naturally occurring peptides at up to 30% of amino acid positions by up to 1, 2, 3, 4, 5, 6, 7, 8, 9 or 10 position changes. Each deletion or substitution of a natural amino acid residue is considered a position change as is the insertion of a residue without substitution. Amino acids substitutions are often conservative substitutions.

[0038] Unless otherwise indicated, reference to A β fragments includes fragments of the natural human amino acid sequences indicated above as well as analogs including allelic, species and induced variants. Analogs of A β fragments induce antibodies that specifically bind with a natural A β peptide (e.g., A β 42). Analogs of A β fragments typically differ from naturally occurring peptide fragment at up to about 30% of amino acid positions. For example, an analog of A β 15-21 may vary by up to 1, 2, 3 or 4 10 position changes. Each deletion or substitution of a natural amino acid residue is considered a position change as is the insertion of a residue without substitution. Amino acids substitutions are often conservative substitutions.

[0039] Some analogs of Aβ or Aβ fragments also include unnatural amino acids or modifications of N or C terminal amino acids at a one, two, five, ten or even all positions. For example, the natural aspartic acid residue at position 1 and/or 7 of Aβ can be replaced with iso-aspartic acid. Examples of unnatural amino acids are D, alpha, alpha-disubstituted amino acids, N-alkyl amino acids, lactic acid, 4-hydroxyproline, gamma-carboxyglutamate, epsilon-N,N,N-trimethyllysine, epsilon-N-acetyllysine, O-phosphoserine, N-acetylserine, N-formylmethionine, 3-methylhistidine, 5-hydroxylysine, omega-N-methylarginine, β-alanine, ornithine, norleucine, norvaline, hydroxproline, thyroxine, gamma-amino butyric acid, homoserine, citrulline, and isoaspartic acid. Some therapeutic agents of the invention are all-D peptides, e.g., all-D Aβ or all-D Aβ fragment, and all-D peptide analogs. Fragments and analogs can be screened for prophylactic or therapeutic efficacy in transgenic animal models in comparison with untreated or placebo controls as described below.

[0040] $A\beta$, its fragments, and analogs can be synthesized by solid phase peptide synthesis or recombinant expression, or can be obtained from natural sources. Automatic peptide synthesizers are commercially available from numerous suppliers, such as Applied

Biosystems, Foster City, California. Recombinant expression can be in bacteria, such as E. coli, yeast, insect cells or mammalian cells. Procedures for recombinant expression are described by Sambrook et al., *Molecular Cloning: A Laboratory Manual* (C.S.H.P. Press, NY 2d ed., 1989). Some forms of Aβ peptide are also available commercially (e.g., American Peptides Company, Inc., Sunnyvale, CA and California Peptide Research, Inc. Napa, CA).

[0041] Therapeutic agents also include longer polypeptides that include, for example, an immunogenic fragment of A β peptide, together with one or more other amino acids flanking the A β peptide one or one or both sides. For example, preferred agents include fusion proteins comprising a segment of A β fused to a heterologous amino acid sequence that induces a helper T-cell response against the heterologous amino acid sequence and thereby a B-cell response against the A β segment. One or more flanking heterologous amino acids can also be used to cap an A β peptide to protect it from degradation in manufacture, storage or use. Such polypeptides can be screened for prophylactic or therapeutic efficacy in animal models in comparison with untreated or placebo controls as described below. Therapeutic agents of the invention include an immunogenic fragment of A β flanked by polylysine sequences. The polylysine sequences can be fused to the N-terminus, the C terminus, or both the N- and C-terminus of A β or an immunogenic fragment of A β . The A β peptide, analog, active fragment or other polypeptide can be administered in associated or multimeric form or in dissociated form Therapeutic agents also include multimers of monomeric immunogenic agents.

[0042] In a further variation, an immunogenic fragment of Aβ can be presented by a virus or a bacteria as part of an immunogenic composition. A nucleic acid encoding the immunogenic peptide is incorporated into a genome or episome of the virus or bacteria. Optionally, the nucleic acid is incorporated in such a manner that the immunogenic peptide is expressed as a secreted protein or as a fusion protein with an outer surface protein of a virus or a transmembrane protein of a bacteria so that the peptide is displayed. Viruses or bacteria used in such methods should be nonpathogenic or attenuated. Suitable viruses include adenovirus, HSV, Venezuelan equine encephalitis virus and other alpha viruses, vesicular stomatitis virus, and other rhabdo viruses, vaccinia and fowl pox. Suitable bacteria include Salmonella and Shigella. Fusion of an immunogenic peptide to HBsAg of HBV is particularly suitable.

[0043] Therapeutic agents also include peptides and other compounds that do not necessarily have a significant amino acid sequence similarity with $A\beta$ but nevertheless serve as mimetics of $A\beta$ and induce a similar immune response. For example, any peptides and proteins forming β -pleated sheets can be screened for suitability. Anti-idiotypic antibodies against monoclonal antibodies to $A\beta$ or other amyloidogenic peptides can also be used. Such anti-Id antibodies mimic the antigen and generate an immune response to it (see *Essential Immunology* (Roit ed., Blackwell Scientific Publications, Palo Alto, 6th ed.), p. 181). Agents other than $A\beta$ peptides should induce an immunogenic response against one or more of the preferred segments of $A\beta$ listed above (e.g., 15-24). Preferably, such agents induce an immunogenic response that is specifically directed to one of these segments without being directed to other segments of $A\beta$.

[0044] Random libraries of peptides or other compounds can also be screened for suitability. Combinatorial libraries can be produced for many types of compounds that can be synthesized in a step-by-step fashion. Such compounds include polypeptides, beta-turn mimetics, polysaccharides, phospholipids, hormones, prostaglandins, steroids, aromatic compounds, heterocyclic compounds, benzodiazepines, oligomeric N-substituted glycines and oligocarbamates. Large combinatorial libraries of the compounds can be constructed by the encoded synthetic libraries (ESL) method described in Affymax, WO 95/12608, Affymax, WO 93/06121, Columbia University, WO 94/08051, Pharmacopeia, WO 95/35503 and Scripps, WO 95/30642 (each of which is incorporated by reference for all purposes). Peptide libraries can also be generated by phage display methods. See, *e.g.*, Devlin, WO 91/18980.

[0045] Combinatorial libraries and other compounds are initially screened for suitability by determining their capacity to specifically bind to antibodies or lymphocytes (B or T) known to be specific for $A\beta$ or other amyloidogenic peptides. For example, initial screens can be performed with any polyclonal sera or monoclonal antibody to $A\beta$ or a fragment thereof. Compounds can then be screened for specifically binding to a specific epitope within $A\beta$ (e.g., 15-24). Compounds can be tested by the same procedures described for mapping antibody epitope specificities. Compounds identified by such screens are then further analyzed for capacity to induce antibodies or reactive lymphocytes to $A\beta$ or fragments thereof. For example, multiple dilutions of sera can be tested on microtiter plates that have been precoated with $A\beta$ or a fragment thereof and a standard ELISA can be performed to test

for reactive antibodies to Aβ or the fragment. Compounds can then be tested for prophylactic and therapeutic efficacy in transgenic animals predisposed to an amyloidogenic disease, as described in the Examples. Such animals include, for example, mice bearing a 717 mutation of APP described by Games et al., supra, and mice bearing a 670/671 Swedish mutation of APP such as described by McConlogue et al., US 5,612,486 and Hsiao et al., Science, 274, 99 (1996); Staufenbiel et al., Proc. Natl. Acad. Sci. USA, 94:13287-13292 (1997); Sturchler-Pierrat et al., Proc. Natl. Acad. Sci. USA, 94:13287-13292 (1997); Borchelt et al., Neuron, 19:939-945 (1997)). The same screening approach can be used on other potential agents analogs of Aβ and longer peptides including fragments of Aβ, described above.

IV. Conjugates

Some agents for inducing an immune response contain the appropriate epitope for inducing an immune response against LBs but are too small to be immunogenic. In this situation, a peptide immunogen can be linked to a suitable carrier molecule to form a conjugate which helps elicit an immune response. A single agent can be linked to a single carrier, multiple copies of an agent can be linked to multiple copies of a carrier, which are in turn linked to each other, multiple copies of an agent can be linked to a single copy of a carrier, or a single copy of an agent can be linked to multiple copies of a carrier, or different carriers. Suitable carriers include serum albumins, keyhole limpet hemocyanin. immunoglobulin molecules, thyroglobulin, ovalbumin, tetanus toxoid, or a toxoid from other pathogenic bacteria, such as diphtheria, E. coli, cholera, or H. pylori, or an attenuated toxin derivative. T cell epitopes are also suitable carrier molecules. Some conjugates can be formed by linking agents of the invention to an immunostimulatory polymer molecule (e.g., tripalmitoyl-S-glycerine cysteine (Pam₃Cys), mannan (a manose polymer), or glucan (a beta 1→2 polymer)), cytokines (e.g., IL-1, IL-1 alpha and beta peptides, IL-2, gamma-INF, IL-10, GM-CSF), and chemokines (e.g., MIP1alpha and beta, and RANTES). Immunogenic agents can also be linked to peptides that enhance transport across tissues, as described in O'Mahony, WO 97/17613 and WO 97/17614. Immunogens may be linked to the carries with or with out spacers amino acids (e.g., gly-gly).

[0047] Some conjugates can be formed by linking agents of the invention to at least one T cell epitope. Some T cell epitopes are promiscuous while other T cell epitopes are universal. Promiscuous T cell epitopes are capable of enhancing the induction of T cell immunity in a wide variety of subjects displaying various HLA types. In contrast to promiscuous T cell

epitopes, universal T cell epitopes are capable of enhancing the induction of T cell immunity in a large percentage, e.g., at least 75%, of subjects displaying various HLA molecules encoded by different HLA-DR alleles.

[0048] A large number of naturally occurring T-cell epitopes exist, such as, tetanus toxoid (e.g., the P2 and P30 epitopes), Hepatitis B surface antigen, pertussis, toxoid, measles virus F protein, Chlamydia trachomitis major outer membrane protein, diphtheria toxoid, Plasmodium falciparum circumsporozite T, Plasmodium falciparum CS antigen, Schistosoma mansoni triose phosphate isomersae, Escherichia coli TraT, and Influenza virus hemagluttinin (HA). The immunogenic peptides of the invention can also be conjugated to the T-cell epitopes described in Sinigaglia F. et al., Nature, 336:778-780 (1988); Chicz R.M. et al., J. Exp. Med., 178:27-47 (1993); Hammer J. et al., Cell 74:197-203 (1993); Falk K. et al., Immunogenetics, 39:230-242 (1994); WO 98/23635; and, Southwood S. et al. J. Immunology, 160:3363-3373 (1998) (each of which is incorporated herein by reference for all purposes). Further examples include:

Influenza Hemagluttinin: HA₃₀₇₋₃₁₉

Malaria CS: T3 epitope EKKIAKMEKASSVFNV (SEQ ID NO:4)

Hepatitis B surface antigen: HBsAg₁₉₋₂₈ FFLLTRILTI (SEQ ID NO:5)

Heat Shock Protein 65: hsp65₁₅₃₋₁₇₁ DQSIGDLIAEAMDKVGNEG (SEQ ID NO:6)

bacille Calmette-Guerin QVHFQPLPPAVVKL (SEQ ID NO:7)

Tetanus toxoid: TT₈₃₀₋₈₄₄ QYIKANSKFIGITEL (SEQ ID NO:8)

Tetanus toxoid: TT₉₄₇₋₉₆₇ FNNFTVSFWLRVPKVSASHLE (SEQ ID NO:9)

HIV gp120 T1: KQIINMWQEVGKAMYA (SEQ ID NO:10).

[0049] Alternatively, the conjugates can be formed by linking agents of the invention to at least one artificial T-cell epitope capable of binding a large proportion of MHC Class II molecules., such as the pan DR epitope ("PADRE"). PADRE is described in US 5,736141, WO 95/07707, and Alexander J et al., Immunity, 1:751-761 (1994) (each of which is incorporated herein by reference for all purposes). A preferred PADRE peptide is AKXVAAWTLKAAA (SEQ ID NO:11), (common residues bolded) wherein X is preferably cyclohexylalanine tyrosine or phenylalanine, with cyclohexylalanine being most preferred.

[0050] Immunogenic agents can be linked to carriers by chemical crosslinking. Techniques for linking an immunogen to a carrier include the formation of disulfide linkages using N-succinimidyl-3-(2-pyridyl-thio) propionate (SPDP) and succinimidyl 4-(N-maleimidomethyl)cyclohexane-1-carboxylate (SMCC) (if the peptide lacks a sulfhydryl group, this can be provided by addition of a cysteine residue). These reagents create a disulfide linkage between themselves and peptide cysteine resides on one protein and an amide linkage through the epsilon-amino on a lysine, or other free amino group in other amino acids. A variety of such disulfide/amide-forming agents are described by *Immun. Rev.* 62, 185 (1982). Other bifunctional coupling agents form a thioether rather than a disulfide linkage. Many of these thio-ether-forming agents are commercially available and include reactive esters of 6-maleimidocaproic acid, 2-bromoacetic acid, and 2-iodoacetic acid, 4-(N-maleimido-methyl)cyclohexane-1-carboxylic acid. The carboxyl groups can be activated by combining them with succinimide or 1-hydroxyl-2-nitro-4-sulfonic acid, sodium salt.

[0051] Immunogenicity can be improved through the addition of spacer residues (e.g., Gly-Gly) between the T_h epitope and the peptide immunogen of the invention. In addition to physically separating the T_h epitope from the B cell epitope (i.e., the peptide immunogen), the glycine residues can disrupt any artificial secondary structures created by the joining of the T_h epitope with the peptide immunogen, and thereby eliminate interference between the T and/or B cell responses. The conformational separation between the helper epitope and the antibody eliciting domain thus permits more efficient interactions between the presented immunogen and the appropriate T_h and B cells.

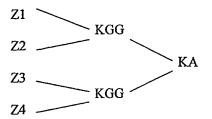
[0052] To enhance the induction of T cell immunity in a large percentage of subjects displaying various HLA types to an agent of the present invention, a mixture of conjugates with different T_h cell epitopes can be prepared. The mixture may contain a mixture of at least two conjugates with different T_h cell epitopes, a mixture of at least three conjugates with different T_h cell epitopes, or a mixture of at least four conjugates with different T_h cell epitopes. The mixture may be administered with an adjuvant.

[0053] Immunogenic peptides can also be expressed as fusion proteins with carriers (i.e., heterologous peptides). The immunogenic peptide can be linked at its amino terminus, its carboxyl terminus, or both to a carrier. Optionally, multiple repeats of the immunogenic peptide can be present in the fusion protein. Optionally, an immunogenic peptide can be linked to multiple copies of a heterologous peptide, for example, at both the N and C termini

of the peptide. Optionally, multiple copies of an immunogenic peptide can be linked to multiple copies of a heterologous peptide, which are linked to each other. Some carrier peptides serve to induce a helper T-cell response against the carrier peptide. The induced helper T-cells in turn induce a B-cell response against the immunogenic peptide linked to the carrier.

[0054] Some examples of fusion proteins suitable for use in the invention are shown below. Some of these fusion proteins comprise segments of A β linked to tetanus toxoid epitopes such as described in US 5,196,512, EP 378,881 and EP 427,347. Some fusion proteins comprise segments of A β linked to at least one PADRE peptide described in US 5,736,142. Some heterologous peptides are promiscuous T-cell epitopes, while other heterologous peptides are universal T-cell epitopes. In some methods, the agent for administration is simply a single fusion protein with an A β segment linked to a heterologous segment in linear configuration. The therapeutic agents of the invention can be represented using a formula. For example, in some methods, the agent is multimer of fusion proteins represented by the formula 2^x , in which x is an integer from 1-5. Preferably x is 1, 2 or 3, with 2 being most preferred. When x is two, such a multimer has four fusion proteins linked in a preferred configuration referred to as MAP4 (see US 5,229,490).

[0055] The MAP4 configuration is shown below, where branched structures are produced by initiating peptide synthesis at both the N terminal and side chain amines of lysine. Depending upon the number of times lysine is incorporated into the sequence and allowed to branch, the resulting structure will present multiple N termini. In this example, four identical N termini have been produced on the branched lysine-containing core. Such multiplicity greatly enhances the responsiveness of cognate B cells. In the examples below, Z refers to an immunogenic fragment of $A\beta$, and Z1-4 refer to immunogenic fragment(s) of $A\beta$. The fragments can be the same as each other or different.



[0056] Other examples of fusion proteins include:

Z-Tetanus toxoid 830-844 in a MAP4 configuration:

Z-QYIKANSKFIGITEL (SEQ ID NO:12)

Z-Tetanus toxoid 947-967 in a MAP4 configuration:

Z-FNNFTVSFWLRVPKVSASHLE (SEQ ID NO:13)

Z-Tetanus toxoid 830-844 in a MAP4 configuration:

Z-QYIKANSKFIGITEL (SEQ ID NO:14)

Z-Tetanus toxoid 830-844 + 947-967 in a linear configuration:

Z-QYIKANSKFIGITELFNNFTVSFWLRVPKVSASHLE (SEQ ID NO:15)

PADRE peptide (all in linear configurations), wherein X is preferably cyclohexylalanine, tyrosine or phenylalanine, with cyclohexylalanine being most preferred-Z:

AKXVAAWTLKAAA-Z (SEQ ID NO:16)

Z x 3-PADRE peptide:

Z-Z-Z-AKXVAAWTLKAAA (SEQ ID NO:17)

Z - ovalbumin 323-339 in a linear configuration:

Z-ISQAVHAAHAEINEAGR (SEQ ID NO:20)

Further examples of fusion proteins include:

AKXVAAWTLKAAA-Z-Z-Z-Z (SEQ ID NO:18)

Z-AKXVAAWTLKAAA (SEQ ID NO:19)

PKYVKQNTLKLAT-Z-Z-Z (SEQ ID NO:21)

Z-PKYVKQNTLKLAT-Z (SEQ ID NO:22)

Z-Z-Z-PKYVKQNTLKLAT (SEQ ID NO:23)

Z-Z-PKYVKQNTLKLAT (SEQ ID NO:24)

Z-PKYVKQNTLKLAT-EKKIAKMEKASSVFNV-QYIKANSKFIGITEL-

FNNFTVSFWLRVPKVSASHLE-Z-

Z-Z-Z-QYIKANSKFIGITEL-FNNFTVSFWLRVPKVSASHLE (SEQ ID NO:25)

Z-QYIKANSKFIGITELCFNNFTVSFWLRVPKVSASHLE-Z-

QYIKANSKFIGITELCFNNFTVSFWLRVPKVSASHLE-Z (SEQ ID NO:26)

Z-QYIKANSKFIGITEL (SEQ ID NO:27) on a 2 branched resin: fragments can be the same as each other or different.

[0057] The same or similar carrier proteins and methods of linkage can be used for generating immunogens to be used in generation of antibodies against $A\beta$ or an immunogenic fragment of $A\beta$. For example, $A\beta$ or an immunogenic fragment of $A\beta$ linked to a carrier can be administered to a laboratory animal in the production of monoclonal antibodies to $A\beta$ or an immunogenic fragment of $A\beta$.

V. Nucleic Acid Encoding Therapeutic Agents

[0058] Immune responses against amyloid deposits can also be induced by administration of nucleic acids encoding segments of Aβ peptide, and fragments thereof, other peptide immunogens, or antibodies and their component chains used for passive immunization. Such nucleic acids can be DNA or RNA. A nucleic acid segment encoding an immunogen is typically linked to regulatory elements, such as a promoter and enhancer, that allow expression of the DNA segment in the intended target cells of a patient. For expression in blood cells, as is desirable for induction of an immune response, promoter and enhancer elements from light or heavy chain immunoglobulin genes or the CMV major intermediate early promoter and enhancer are suitable to direct expression. The linked regulatory elements and coding sequences are often cloned into a vector. For administration of double-chain antibodies, the two chains can be cloned in the same or separate vectors. The nucleic acids encoding therapeutic agents of the invention can also encode at least one T cell epitope. The disclosures herein which relate to the use of adjuvants and the use of carriers apply mutatis mutandis to their use with the nucleic acids encoding the therapeutic agents of the present invention.

[0059] A number of viral vector systems are available including retroviral systems (see, e.g., Lawrie and Tumin, Cur. Opin. Genet. Develop. 3, 102-109 (1993)); adenoviral vectors (see, e.g., Bett et al., J. Virol. 67, 5911 (1993)); adeno-associated virus vectors (see, e.g., Zhou et al., J. Exp. Med. 179, 1867 (1994)), viral vectors from the pox family including vaccinia virus and the avian pox viruses, viral vectors from the alpha virus genus such as those derived from Sindbis and Semliki Forest Viruses (see, e.g., Dubensky et al., J. Virol.

70, 508-519 (1996)), Venezuelan equine encephalitis virus (see US 5,643,576) and rhabdoviruses, such as vesicular stomatitis virus (see WO 96/34625) and papillomaviruses (Ohe et al., *Human Gene Therapy* 6, 325-333 (1995); Woo et al., WO 94/12629 and Xiao & Brandsma, *Nucleic Acids. Res.* 24, 2630-2622 (1996)).

[0060] DNA encoding an immunogen, or a vector containing the same, can be packaged into liposomes. Suitable lipids and related analogs are described by US 5,208,036, 5,264,618, 5,279,833 and 5,283,185. Vectors and DNA encoding an immunogen can also be adsorbed to or associated with particulate carriers, examples of which include polymethyl methacrylate polymers and polylactides and poly(lactide-co-glycolides), see, e.g., McGee et al., J. Micro Encap. (1996).

[0061] Gene therapy vectors or naked DNA can be delivered in vivo by administration to an individual patient, typically by systemic administration (e.g., intravenous, intraperitoneal, nasal, gastric, intradermal, intramuscular, subdermal, or intracranial infusion) or topical application (see e.g., US 5,399,346). Such vectors can further include facilitating agents such as bupivacine (US 5,593,970). DNA can also be administered using a gene gun. (See Xiao & Brandsma, supra.) The DNA encoding an immunogen is precipitated onto the surface of microscopic metal beads. The microprojectiles are accelerated with a shock wave or expanding helium gas, and penetrate tissues to a depth of several cell layers. For example, The Accel™ Gene Delivery Device manufactured by Agacetus, Inc. Middleton WI is suitable. Alternatively, naked DNA can pass through skin into the blood stream simply by spotting the DNA onto skin with chemical or mechanical irritation (see WO 95/05853).

[0062] In a further variation, vectors encoding immunogens can be delivered to cells ex vivo, such as cells explanted from an individual patient (e.g., lymphocytes, bone marrow aspirates, tissue biopsy) or universal donor hematopoietic stem cells, followed by reimplantation of the cells into a patient, usually after selection for cells which have incorporated the vector.

VI. Adjuvants

[0063] Immunogenic agents of the invention, such as peptides, are sometimes administered in combination with an adjuvant. The adjuvant increases the titer of induced antibodies and/or the binding affinity of induced antibodies relative to the situation if the peptide were used alone. A variety of adjuvants can be used in combination with an immunogenic fragment of $A\beta$, to elicit an immune response. Preferred adjuvants augment the intrinsic

response to an immunogen without causing conformational changes in the immunogen that affect the qualitative form of the response. Preferred adjuvants include aluminum hydroxide and aluminum phosphate, 3 De-O-acylated monophosphoryl lipid A (MPLTM) (see GB 2220211 (RIBI ImmunoChem Research Inc., Hamilton, Montana, now part of Corixa). StimulonTM QS-21 is a triterpene glycoside or saponin isolated from the bark of the Quillaja Saponaria Molina tree found in South America (see Kensil et al., in Vaccine Design: The Subunit and Adjuvant Approach (eds. Powell & Newman, Plenum Press, NY, 1995); US Patent No. 5,057,540), (Aquila BioPharmaceuticals, Framingham, MA). Other adjuvants are oil in water emulsions (such as squalene or peanut oil), optionally in combination with immune stimulants, such as monophosphoryl lipid A (see Stoute et al., N. Engl. J. Med. 336, 86-91 (1997)), pluronic polymers, and killed mycobacteria. Another adjuvant is CpG (WO 98/40100). Adjuvants can be administered as a component of a therapeutic composition with an active agent or can be administered separately, before, concurrently with, or after administration of the therapeutic agent.

A preferred class of adjuvants is aluminum salts (alum), such as alum hydroxide. alum phosphate, alum sulfate. Such adjuvants can be used with or without other specific immunostimulating agents such as MPL or 3-DMP, QS-21, polymeric or monomeric amino acids such as polyglutamic acid or polylysine. Another class of adjuvants is oil-in-water emulsion formulations. Such adjuvants can be used with or without other specific immunostimulating agents such as muramyl peptides (e.g., N-acetylmuramyl-L-threonyl-Disoglutamine (thr-MDP), N-acetyl-normuramyl-L-alanyl-D-isoglutamine (nor-MDP), Nacetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'dipalmitoyl-sn-glycero-3hydroxyphosphoryloxy)-ethylamine (MTP-PE), N-acetylglucsaminyl-N-acetylmuramyl-L-Al-D-isoglu-L-Ala-dipalmitoxy propylamide (DTP-DPP) theramideTM), or other bacterial cell wall components. Oil-in-water emulsions include (a) MF59 (WO 90/14837), containing 5% Squalene, 0.5% Tween 80, and 0.5% Span 85 (optionally containing various amounts of MTP-PE) formulated into submicron particles using a microfluidizer such as Model 110Y microfluidizer (Microfluidics, Newton MA), (b) SAF, containing 10% Squalene, 0.4% Tween 80, 5% pluronic-blocked polymer L121, and thr-MDP, either microfluidized into a submicron emulsion or vortexed to generate a larger particle size emulsion, and (c) Ribi™ adjuvant system (RAS), (Ribi ImmunoChem, Hamilton, MT) containing 2% squalene, 0.2% Tween 80, and one or more bacterial cell wall components from the group consisting of

monophosphoryllipid A (MPL), trehalose dimycolate (TDM), and cell wall skeleton (CWS), preferably MPL + CWS (DetoxTM).

[0065] Another class of preferred adjuvants is saponin adjuvants, such as Stimulon™ (QS-21, Aquila, Framingham, MA) or particles generated therefrom such as ISCOMs (immunostimulating complexes) and ISCOMATRIX. Other adjuvants include RC-529, GM-CSF and Complete Freund's Adjuvant (CFA) and Incomplete Freund's Adjuvant (IFA). Other adjuvants include cytokines, such as interleukins (e.g., IL-1 α and β peptides,, IL-2, IL-4, IL-6, IL-12, IL13, and IL-15), macrophage colony stimulating factor (M-CSF), granulocyte-macrophage colony stimulating factor (GM-CSF), tumor necrosis factor (TNF), chemokines, such as MIP1α and β and RANTES. Another class of adjuvants is glycolipid analogues including N-glycosylamides, N-glycosylureas and N-glycosylcarbamates, each of which is substituted in the sugar residue by an amino acid, as immuno-modulators or adjuvants (see US Pat. No. 4,855,283). Heat shock proteins, e.g., HSP70 and HSP90, may also be used as adjuvants.

[0066] An adjuvant can be administered with an immunogen as a single composition, or can be administered before, concurrent with or after administration of the immunogen. Immunogen and adjuvant can be packaged and supplied in the same vial or can be packaged in separate vials and mixed before use. Immunogen and adjuvant are typically packaged with a label indicating the intended therapeutic application. If immunogen and adjuvant are packaged separately, the packaging typically includes instructions for mixing before use. The choice of an adjuvant and/or carrier depends on the stability of the immunogenic formulation containing the adjuvant, the route of administration, the dosing schedule, the efficacy of the adjuvant for the species being vaccinated, and, in humans, a pharmaceutically acceptable adjuvant is one that has been approved or is approvable for human administration by pertinent regulatory bodies. For example, Complete Freund's adjuvant is not suitable for human administration. Alum, MPL and QS-21 are preferred. Optionally, two or more different adjuvants can be used simultaneously. Preferred combinations include alum with MPL, alum with QS-21, MPL with QS-21, MPL or RC-529 with GM-CSF, and alum, QS-21 and MPL together. Also, Incomplete Freund's adjuvant can be used (Chang et al., Advanced Drug Delivery Reviews 32, 173-186 (1998)), optionally in combination with any of alum, QS-21, and MPL and all combinations thereof.

VII. Passive Administration Of Antibodies

[0067] Active immunization with fragments of $A\beta$ can be combined with passive administration of antibodies. The antibodies used for passive administration can be antibodies to N-terminal epitopes of $A\beta$ for induction of a phagocytic clearing response of plaques, or can be antibodies to central or C-terminal regions of $A\beta$ for clearing soluble $A\beta$. In some methods, passive administration with an antibody to an N-terminal region antibody is performed first to clear existing amyloid deposits. Subsequently, a fragment from a central or C-terminal region of $A\beta$ is administered to prevent further deposition of amyloid deposits from soluble $A\beta$. In other method, active administration with a fragment to a central or C-terminal portion of $A\beta$ is performed first to generate antibodies that clear soluble $A\beta$. Then when the level of antibodies in the blood starts to wane, an additional dose is supplied by passive administration of antibodies that specifically bind to a central or C-terminal epitope of $A\beta$.

Antibodies suitable for use in passive administration are described in WO 00/72880 and WO 02/46237 incorporated by reference. Preferred antibodies specifically binding to an N-terminal epitope of Aβ bind to an epitope starting at resides 1-3 and ending at residues 7-11 of Aβ. Some preferred antibodies specifically bind to epitopes within amino acids 1-3, 1-4, 1-5, 1-6, 1-7 or 3-7. Some preferred antibodies specifically bind to an epitope starting at resides 1-3 and ending at residues 7-11 of Aβ. Such antibodies typically specifically bind to amyloid deposits but may or may not bind to soluble Aβ. Some preferred antibodies specifically binding to a C-terminal epitope of Aβ specifically bind to a naturally occurring long form of Aβ (i.e., Aβ42 and Aβ43 without specifically binding to a naturally occurring short form of Aß (i.e., Aß39, 40 or 41). Antibodies to C-terminal and central epitopes of typically specifically bind to soluble without specific binding to amyloid deposits. When an antibody is said to specifically bind to an epitope within specified residues, such as Aß 1-5 for example, what is meant is that the antibody specifically binds to a polypeptide containing the specified residues (i.e., A\beta 1-5 in this an example). Such an antibody does not necessarily contact every residue within A\beta 1-5. Nor does every single amino acid substitution or deletion with in A\u00e31-5 necessarily significantly affect binding affinity.

Epitope specificity of an antibody can be determined, for example, as described by WO 00/72880.

[0069] Antibodies can be polyclonal or monoclonal. Polyclonal sera typically contain mixed populations of antibodies specifically binding to several epitopes along the length of Aβ. However, polyclonal sera can be specific to a particular segment of Aβ, such as Aβ1-10. Preferred antibodies are chimeric, humanized (see Queen et al., *Proc. Natl. Acad. Sci. USA* 86:10029-10033 (1989) and WO 90/07861, US 5,693,762, US 5,693,761, US 5,585,089, US 5,530,101 and Winter, US 5,225,539), or human (Lonberg et al., WO93/12227 (1993); US 5,877,397, US 5,874,299, US 5,814,318, US 5,789,650, US 5,770,429, US 5,661,016, US 5,633,425, US 5,625,126, US 5,569,825, US 5,545,806, *Nature* 148, 1547-1553 (1994), *Nature Biotechnology* 14, 826 (1996), Kucherlapati, WO 91/10741 (1991)). Several mouse antibodies of different binding specificities are available as starting materials for making humanized antibodies. Human isotype IgG1 is preferred for antibodies to the N-terminal region of because of it having highest affinity of human isotypes for the FcRI receptor on phagocytic cells. Some antibodies specifically bind to Aβ with a binding affinity greater than or equal to about 10⁷, 10⁸, 10⁹, or 10¹⁰ M⁻¹.

VIII. Patients Amenable To Treatment

[0070] Patients amenable to treatment include individuals at risk of disease but not showing symptoms, as well as patients presently showing symptoms. In the case of Alzheimer's disease, virtually anyone is at risk of suffering from Alzheimer's disease if he or she lives long enough. Therefore, the present methods can be administered prophylactically to the general population without the need for any assessment of the risk of the subject patient. The present methods are especially useful for individuals who do have a known genetic risk of Alzheimer's disease. Such individuals include those having relatives who have experienced this disease, and those whose risk is determined by analysis of genetic or biochemical markers. Genetic markers of risk toward Alzheimer's disease include mutations in the APP gene, particularly mutations at position 717 and positions 670 and 671 referred to as the Hardy and Swedish mutations respectively (see Hardy, TINS, supra). Other markers of risk are mutations in the presenilin genes, PS1 and PS2, and ApoE4, family history of AD, hypercholesterolemia or atherosclerosis. Individuals presently suffering from Alzheimer's disease can be recognized from characteristic dementia, as well as the presence of risk factors described above. In addition, a number of diagnostic tests are available for identifying

individuals who have AD. These include measurement of CSF tau and A β 42 levels. Elevated tau and decreased A β 42 levels signify the presence of AD. Individuals suffering from Alzheimer's disease can also be diagnosed by ADRDA criteria as discussed in WO 00/72880.

[0071] In asymptomatic patients, treatment can begin at any age (e.g., 10, 20, 30). Usually, however, it is not necessary to begin treatment until a patient reaches 40, 50, 60 or 70. Treatment typically entails multiple dosages over a period of time. Treatment can be monitored by assaying antibody, or activated T-cell (a side effect) or B-cell responses to the therapeutic agent (e.g., Aß peptide) over time. If the response falls, a booster dosage is indicated. In the case of potential Down's syndrome patients, treatment can begin antenatally by administering therapeutic agent to the mother or shortly after birth.

IX. <u>Treatment Regimes</u>

[0072] In general treatment regimes involve administering an agent effective to induce an immunogenic response to A\beta, preferably an immunogenic fragment of A\beta to a patient. In prophylactic applications, pharmaceutical compositions or medicaments are administered to a patient susceptible to, or otherwise at risk of, Alzheimer's disease in an amount sufficient to eliminate or reduce the risk, lessen the severity, or delay the onset of the disease, including physiological, biochemical, histologic and/or behavioral symptoms of the disease, its complications and intermediate pathological phenotypes presenting during development of the disease. In therapeutic applications, an agent is administered to a patient suspected of, or already suffering from such a disease in a regime comprising an amount and frequency of administration of the agent sufficient to cure, or at least partially arrest, or inhibit deterioration of the symptoms of the disease (physiological, biochemical, histologic and/or behavioral), including its complications and intermediate pathological phenotypes in development of the disease. In some methods, administration of agent reduces or eliminates myocognitive impairment in patients that have not yet developed characteristic Alzheimer's pathology. An amount adequate to accomplish therapeutic or prophylactic treatment is defined as a therapeutically- or prophylactically-effective dose. A combination of amount and dosage frequency adequate to accomplish the therapeutic or prophylactic treatment is defined as a therapeutically- or prophylactically-effective regime. In both prophylactic and therapeutic regimes, agents are usually administered in several dosages until a sufficient immune response has been achieved. A dosage and frequency of administrations adequate to

accomplish therapeutic or prophylactic treatment is defined as a therapeutically- or prophylactically-effective regime. Typically, the patient's immune response is monitored and repeated dosages are given if the immune response starts to wane. The immune response can be monitored by detecting antibodies to AB in the blood in the patient, detecting levels of AB or plaques in the brain or symptoms by a psychometric measure, such as the MMSE, and the ADAS, which is a comprehensive scale for evaluating patients with Alzheimer's Disease status and function.

Effective doses of the agents and compositions of the present invention, for the treatment of the above described conditions vary depending upon many different factors, including means of administration, target site, physiological state of the patient, whether the patient is human or an animal, other medications administered, and whether treatment is prophylactic or therapeutic. Usually, the patient is a human but nonhuman mammals including transgenic mammals can also be treated. Treatment dosages need to be titrated to optimize safety and efficacy. The amount of immunogen depends on whether adjuvant is also administered, with higher dosages being required in the absence of adjuvant. The amount of an immunogen for administration sometimes varies from 1-500 µg per patient and more usually from 5-500 µg per injection for human administration. Occasionally, a higher dose of 1-2 mg per injection is used. Typically at least 10, 20, 50 or 100 µg is used for each human injection. The mass of immunogen also depends on the mass ratio of immunogenic epitope within the immunogen to the mass of immunogen as a whole. Typically, 10⁻³ to 10⁻⁵ micromoles of immunogenic epitope are used for microgram of immunogen. The timing of injections can vary significantly from once a day, to once a year, to once a decade. On any given day that a dosage of immunogen is given, the dosage is greater than 1 µg/patient and usually greater than 10 µg/ patient if adjuvant is also administered, and greater than 10 μg/patient and usually greater than 100 μg/patient in the absence of adjuvant. A typical regimen consists of an immunization followed by booster injections at time intervals, such as 6 week intervals. Another regimen consists of an immunization followed by booster injections 1, 2 and 12 months later. Another regimen entails an injection every two months for life. Alternatively, booster injections can be on an irregular basis as indicated by monitoring of immune response.

[0074] Doses for nucleic acids encoding immunogens range from about 10 ng to 1 g, 100 ng to 100 mg, 1 µg to 10 mg, or 30-300 µg DNA per patient. Doses for infectious viral vectors vary from 10-100, or more, virions per dose.

[0075] For passive immunization with an antibody (in combination therapies), the dosage ranges from about 0.0001 to 100 mg/kg, and more usually 0.01 to 5 mg/kg, of the host body weight. For example dosages can be 1 mg/kg body weight or 10 mg/kg body weight or within the range of 1-10 mg/kg or in other words, 70 mg or 700 mg or within the range of 70-700 mg, respectively, for a 70 kg patient. An exemplary treatment regime entails administration once per every two weeks or once a month or once every 3 to 6 months. In some methods, two or more monoclonal antibodies with different binding specificities are administered simultaneously, in which case the dosage of each antibody administered falls within the ranges indicated. Antibody is usually administered on multiple occasions. Intervals between single dosages can be weekly, monthly or yearly. Intervals can also be irregular as indicated by measuring blood levels of antibody to AB in the patient. In some methods, dosage is adjusted to achieve a plasma antibody concentration of 1-1000 µg/ml and in some methods 25-300 µg/ml. Alternatively, antibody can be administered as a sustained release formulation, in which case less frequent administration is required. Dosage and frequency vary depending on the half-life of the antibody in the patient. In general, human antibodies show the longest half life, followed by humanized antibodies, chimeric antibodies, and nonhuman antibodies. The dosage and frequency of administration can vary depending on whether the treatment is prophylactic or therapeutic. In prophylactic applications, a relatively low dosage is administered at relatively infrequent intervals over a long period of time. Some patients continue to receive treatment for the rest of their lives. In therapeutic applications, a relatively high dosage at relatively short intervals is sometimes required until progression of the disease is reduced or terminated, and preferably until the patient shows partial or complete amelioration of symptoms of disease. Thereafter, the patent can be administered a prophylactic regime.

[0076] Agents for inducing an immune response can be administered by parenteral, topical, intravenous, oral, subcutaneous, intraarterial, intracranial, intraperitoneal, intranasal or intramuscular means for prophylactic and/or therapeutic treatment. The most typical route of administration of an immunogenic agent is subcutaneous although other routes can be equally effective. The next most common route is intramuscular injection. This type of injection is most typically performed in the arm or leg muscles. In some methods, agents are injected

directly into a particular tissue where deposits have accumulated, e.g., intracranial injection. Intramuscular injection or intravenous infusion are preferred for administration of antibody (in combination therapies). In some methods, particular therapeutic antibodies are injected directly into the cranium. In some methods, antibodies are administered as a sustained release composition or device, such as a MedipadTM device.

[0077] Agents of the invention are often administered as pharmaceutical compositions comprising an active therapeutic agent, i.e., and a variety of other pharmaceutically acceptable components. See Remington's Pharmaceutical Science (15th ed., Mack Publishing Company, Easton, Pennsylvania, 1980). The preferred form depends on the intended mode of administration and therapeutic application. The compositions can also include, depending on the formulation desired, pharmaceutically-acceptable, non-toxic carriers or diluents, which are defined as vehicles commonly used to formulate pharmaceutical compositions for animal or human administration. The diluent is selected so as not to affect the biological activity of the combination. Examples of such diluents are distilled water, physiological phosphate-buffered saline, Ringer's solutions, dextrose solution, and Hank's solution. In addition, the pharmaceutical composition or formulation may also include other carriers, adjuvants, or nontoxic, nontherapeutic, nonimmunogenic stabilizers and the like.

[0078] Pharmaceutical compositions can also include large, slowly metabolized macromolecules such as proteins, polysaccharides such as chitosan, polylactic acids, polyglycolic acids and copolymers (such as latex functionalized sepharose(TM), agarose, cellulose, and the like), polymeric amino acids, amino acid copolymers, and lipid aggregates (such as oil droplets or liposomes). Additionally, these carriers can function as immunostimulating agents (i.e., adjuvants).

[0079] For parenteral administration, agents of the invention can be administered as injectable dosages of a solution or suspension of the substance in a physiologically acceptable diluent with a pharmaceutical carrier that can be a sterile liquid such as water oils, saline, glycerol, or ethanol. Additionally, auxiliary substances, such as wetting or emulsifying agents, surfactants, pH buffering substances and the like can be present in compositions. Other components of pharmaceutical compositions are those of petroleum, animal, vegetable, or synthetic origin, for example, peanut oil, soybean oil, and mineral oil. In general, glycols such as propylene glycol or polyethylene glycol are preferred liquid carriers, particularly for injectable solutions. Antibodies can be administered in the form of a depot injection or

implant preparation which can be formulated in such a manner as to permit a sustained release of the active ingredient. An exemplary composition comprises monoclonal antibody at 5 mg/mL, formulated in aqueous buffer consisting of 50 mM L-histidine, 150 mM NaCl, adjusted to pH 6.0 with HCl. Composition for parenteral administration are typically substantially sterile, isotonic and manufactured under GMP conditions of the FDA or similar body.

[0080] Typically, compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid vehicles prior to injection can also be prepared. The preparation also can be emulsified or encapsulated in liposomes or micro particles such as polylactide, polyglycolide, or copolymer for enhanced adjuvant effect, as discussed above (see Langer, Science 249, 1527 (1990) and Hanes, Advanced Drug Delivery Reviews 28, 97-119 (1997). The agents of this invention can be administered in the form of a depot injection or implant preparation which can be formulated in such a manner as to permit a sustained or pulsatile release of the active ingredient.

[0081] Additional formulations suitable for other modes of administration include oral, intranasal, and pulmonary formulations, suppositories, and transdermal applications.

[0082] For suppositories, binders and carriers include, for example, polyalkylene glycols or triglycerides; such suppositories can be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1%-2%. Oral formulations include excipients, such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, and magnesium carbonate. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain 10%-95% of active ingredient, preferably 25%-70%.

[0083] Topical application can result in transdermal or intradermal delivery. Topical administration can be facilitated by co-administration of the agent with cholera toxin or detoxified derivatives or subunits thereof or other similar bacterial toxins (See Glenn *et al.*, *Nature* 391, 851 (1998)). Co-administration can be achieved by using the components as a mixture or as linked molecules obtained by chemical crosslinking or expression as a fusion protein.

[0084] Alternatively, transdermal delivery can be achieved using a skin path or using transferosomes (Paul et al., Eur. J. Immunol. 25, 3521-24 (1995); Cevc et al., Biochem. Biophys. Acta 1368, 201-15 (1998)).

X. Methods of Monitoring

The invention provides methods of detecting an antibody response against $A\beta$ peptide in a patient suffering from or susceptible to an amyloidogenic disease. The methods are particularly useful for monitoring a course of treatment being administered to a patient. The methods can be used to monitor both therapeutic treatment on symptomatic patients and prophylactic treatment on asymptomatic patients. Some methods entail determining a baseline value of an antibody response in a patient before administering a dosage of an immunogenic agent, and comparing this with a value for the immune response after treatment. A significant increase (i.e., greater than the typical margin of experimental error in repeat measurements of the same sample, expressed as one standard deviation from the mean of such measurements) in value of the antibody response signals a positive treatment outcome (i.e., that administration of the agent has achieved or augmented an immune response). If the value for the antibody response does not change significantly, or decreases, a negative treatment outcome is indicated. In general, patients undergoing an initial course of treatment with an immunogenic agent are expected to show an increase in antibody response with successive dosages, which eventually reaches a plateau. Administration of agent is generally continued while the antibody response is increasing. Attainment of the plateau is an indicator that the administered of treatment can be discontinued or reduced in dosage or frequency.

[0086] In other methods, a control value (i.e., a mean and standard deviation) of an antibody response is determined for a control population. Typically the individuals in the control population have not received prior treatment. Measured values of the antibody response in a patient after administering a therapeutic agent are then compared with the control value. A significant increase relative to the control value (e.g., greater than one standard deviation from the mean) signals a positive treatment outcome. A lack of significant increase or a decrease signals a negative treatment outcome. Administration of agent is generally continued while the antibody response is increasing relative to the control value. As before, attainment of a plateau relative to control values in an indicator that the administration of treatment can be discontinued or reduced in dosage or frequency.

[0087] In other methods, a control value of antibody response (e.g., a mean and standard deviation) is determined from a control population of individuals who have undergone treatment with a therapeutic agent and whose antibody responses have reached a plateau in response to treatment. Measured values of antibody response in a patient are compared with

the control value. If the measured level in a patient is not significantly different (e.g., more than one standard deviation) from the control value, treatment can be discontinued. If the level in a patient is significantly below the control value, continued administration of agent is warranted. If the level in the patient persists below the control value, then a change in treatment regime, for example, use of a different adjuvant, fragment or switch to passive administration may be indicated.

[0088] In other methods, a patient who is not presently receiving treatment but has undergone a previous course of treatment is monitored for antibody response to determine whether a resumption of treatment is required. The measured value of antibody response in the patient can be compared with a value of antibody response previously achieved in the patient after a previous course of treatment. A significant decrease relative to the previous measurement (i.e., greater than a typical margin of error in repeat measurements of the same sample) is an indication that treatment can be resumed. Alternatively, the value measured in a patient can be compared with a control value (mean plus standard deviation) determined in a population of patients after undergoing a course of treatment. Alternatively, the measured value in a patient can be compared with a control value in populations of prophylactically treated patients who remain free of symptoms of disease, or populations of therapeutically treated patients who show amelioration of disease characteristics. In all of these cases, a significant decrease relative to the control level (i.e., more than a standard deviation) is an indicator that treatment should be resumed in a patient.

[0089] The tissue sample for analysis is typically blood, plasma, serum, mucous or cerebrospinal fluid from the patient. The sample is analyzed for indication of an immune response to any form of $A\beta$ peptide, typically $A\beta$ 42 or the peptide used for immunization. The immune response can be determined from the presence of antibodies that specifically bind to $A\beta$ peptide. Antibodies can be detected in a binding assay to a ligand that specifically binds to the antibodies. Typically the ligand is immobilized. Binding can be detected using a labeled anti-idiotypic antibody.

[0090] In combination regimes employing both active and passive administration, analogous approaches can be used to monitor levels of antibody resulting from passive administration as described in WO 00/72880.

Examples

Materials and Methods

[0091] Aβ Fragments. Peptides corresponding to Aβ1-5, Aβ3-9, Aβ5-11, Aβ15-24 and the reverse sequence Aβ5-1 were synthesized contiguous to a 17-amino acid T cell epitope derived from ovalbumin (amino acids 323-339 – ISQAVHAAHAEINEAGR (SEQ ID NO:3)) on a branched peptide framework (triple-lysine core with four peptide arms) to produce a multi-antigen peptide, as described by Tam, J. P. (1988) *Proc. Natl. Acad. Sci. USA* 85, 5409-5413. Polyclonal antibodies (Pab) Aβ1-42 were raised and the immunoglobulin fraction isolated, as previously described by Bard, F. et al., (2000) *Nat. Med.* 6, 916-919. Polyclonal Pab-EL16, Pab-EL17, and Pab-EL20 were obtained from the sera of PDAPP mice immunized with peptides corresponding respectively to Aβ1-7, Aβ15-24, and Aβ3-9 that had been synthesized on a branched framework, as described above. Pab-EL26 was obtained from the sera of mice immunized with Aβ(7-1)-42. The peptides were synthesized by AnaSpec, San Jose, CA, USA.

[0092] Immunization Procedures. 100 μg of A β fragment was administered by intraperitoneal injection in complete Freund's adjuvant, followed by boosts with 100 μg peptide in incomplete Freund's adjuvant at 2 and 4 weeks, and monthly thereafter.

[0093] Antibody Binding to Aggregated and Soluble Aβ1-42. Serum titers (determined by serial dilution) and monoclonal antibody binding to aggregated synthetic Aβ1-42 were performed by ELISA as previously described by Schenk D. et al., (1999) *Nature* 400, 173-177. Soluble Aβ1-42 refers to the synthetic Aβ1-42 peptide sonicated in dimethyl sulfoxide. Serial dilutions of antibody were incubated with 50,000 cpm of ¹²⁵I-Aβ1-42 overnight at room temperature. 50 μl of a slurry containing 75 mg/ml protein A sepharose (Amersham Biosciences, Uppsala, Sweden)/200 μg rabbit anti-mouse IgG (H+L) (Jackson ImmunoResearch, West Grove, PA, USA) was incubated with the diluted antibodies for 1 hr at room temperature, washed twice, and counted on a Wallac gamma counter (PerkinElmer Life Science, Grove, IL, USA). All steps were performed in radioimmunoassay buffer consisting of 10 mM Tris, 0.5 M NaCl, 1 mg/ml gelatin, and 0.5% Nonidet P-40, pH 8.0.

Results

[0094] A series of peptides were compared for their ability to trigger an efficacious antibody response in vivo. Twelve to thirteen month old PDAPP mice were immunized with one of three N-terminal peptide fragments (A\beta 1-5, A\beta 3-9, or A\beta 5-11) or a fragment derived from an internal region of the peptide (AB15-24) (Fig 1a). The internal peptide AB15-24 encompasses the epitope of antibody 266, which exhibits high affinity for soluble AB (Seubert et al., (1992) Nature 359, 325-327.), does not recognize plaques in sections of unfixed AD or PDAPP tissue. Thus, it was of interest to determine whether a polyclonal response directed against this peptide could produce antibodies capable of plaque recognition, or whether reactivity with soluble AB alone was sufficient to provide efficacy. In these studies, a peptide with reverse sequence, AB5-1, served as a negative control. The peptides were synthesized contiguous to a 17-amino acid T cell epitope derived from ovalbumin and were presented in an identical multivalent configuration (see Materials and Methods). All of the peptides (except AB5-1 reversemer) produced sera that recognized aggregated synthetic Aβ1-42 by ELISA, although Aβ5-11 and Aβ15-24 produced significantly higher titers than Aβ1-5 (p<0.01 and p<0.05, respectively) (Fig. 1b). In contrast, only sera against the Nterminal peptides were able to recognize AB within plaques; antisera against AB15-24 did not bind plaques in spite of strong reactivity with the synthetic aggregated peptide (Fig. 1c). There were also differences between the serum groups in their ability to capture soluble $A\beta$ (Fig. 2a). Less than 30% of the sera from mice immunized with A\beta 1-5 or A\beta 3-9 captured the soluble peptide (27% and 5% respectively). In contrast, sera from approximately half of the animals immunized with AB5-11, and all of those immunized with AB15-24, captured soluble Αβ1-42.

[0095] Because the degree of A β deposition can vary greatly as PDAPP mice age, the *in vivo* study was designed with at least 30 animals per group. Efficacy data are shown for individual mice and expressed as the percentage of either amyloid burden or neuritic dystrophy relative to the mean of the control (set at 100%). Immunization with each of the three N-terminal peptides significantly reduced amyloid burden (46-61%, p<0.002) (Fig. 2b). Furthermore, A β 3-9 and A β 5-11significantly reduced neuritic pathology (34% and 41% respectively, p<0.05), (Fig 2c). Immunization with A β 15-24 provided no protection against either amyloid burden or neuritic pathology. These results support plaque binding as one mechanism for antibody efficacy. They also indicate that capture of soluble A β is not

required for reduction of neuritic pathology since the antibody response against A β 3-9 provided strong plaque reactivity and the highest level of protection against neuronal dystrophy, yet exhibited the weakest capacity for recognition of soluble peptide. Antibodies that bind to soluble A β without binding to plaques may also have such activity if administered at higher titers or over longer periods of time. Antibodies that bind to soluble A β without binding to plaques can also be useful in preventing formation and/or further deposition of A β . The high titer of antibodies generated by immunization with A β 15-24 indicates that his fragment and subfragments thereof are particularly useful for generating high titers of soluble antibodies for this purpose.

[0096] Although the foregoing invention has been described in detail for purposes of clarity of understanding, it will be obvious that certain modifications may be practiced within the scope of the appended claims. All publications and patent documents cited herein are hereby incorporated by reference in their entirety for all purposes to the same extent as if each were so individually denoted. Unless otherwise apparent from the context, each element, feature or embodiment of the invention can be used in combination with any other.